# Impact of and correction for instrument sensitivity drift on nanoparticle size measurements by single-particle ICP-MS

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- 12 Abstract: The effect of ICP-MS instrument sensitivity drift on the accuracy of NP size
- 13 measurements using single particle (sp)ICP-MS is investigated. Theoretical modeling and experimental
- 14 measurements of the impact of instrument sensitivity drift are in agreement and indicate that drift can
- 15 impact the measured size of spherical NPs by up to 25 %. Given this substantial bias in the measured
- size, a method was developed using an internal standard to correct for the impact of drift and was
- 17 shown to accurately correct for a decrease in instrument sensitivity of up to 50 % for 30 nm and 60 nm
- 18 gold nanoparticles.
- 19
- 20 Keywords: drift correction, gold nanoparticles, instrument sensitivity drift, internal standard, single-
- 21 particle ICP-MS

### 22 1 Introduction

23 An engineered nanomaterial (ENM) can be defined as a material having any external dimension, 24 internal structure, or surface structure in the nanoscale (approximately 1 nm to 100 nm) and that is 25 designed for a specific purpose or function [1]. The unique and enhanced properties of ENMs compared 26 to traditional materials suggest that they will be used in consumer products increasingly and in more 27 diverse ways in future years. There are many exciting opportunities to employ ENMs in a range of fields, 28 such as composite materials, solar energy, environmental remediation, and aeronautics [2-5]. However, 29 the increased use of ENMs in consumer products and the potential release of ENMs during the product 30 life cycle could have unknown effects on humans and other biological systems, as well as on 31 environmental systems [6-10]. Thus, a substantial research effort is being made to understand the 32 potential environmental, health, and safety (EHS) implications of nanotechnology. One challenging 33 aspect of making measurements for EHS research is the lack of broadly available techniques for 34 accurately and efficiently measuring the size distribution and number concentration of nano-objects 35 (defined as nanomaterials with one, two, or three external dimensions in the nanoscale[1]) in liquid 36 suspension. Techniques such as electron microscopy are time consuming, tedious, and typically require 37 drying the sample prior to analysis, which is a known source of artifacts. Other techniques, such as 38 dynamic light scattering (DLS), are challenged to measure multimodal distributions accurately [11]. NP 39 tracking analysis can overcome the limitations of DLS by detecting small NPs among large ones and 40 determining the number concentration [12], although this method is not element specific and the size 41 distribution of a NP dispersion provided by this technique is sometimes broader than those obtained by 42 other techniques [13]. Nano-objects in suspensions for toxicological tests may undergo changes such as 43 agglomeration/aggregation or dissolution that could lead to inaccurate dosing, if changes to the nano-

44 objects during the test are not measured [14,15].

45 One technique that shows substantial promise for accurately measuring metal containing nano-46 objects suspended in liquid solution is single-particle inductively coupled plasma mass spectrometry 47 (spICP-MS). This technique, first developed by Degueldre et al., [16,17] has recently been used to 48 measure gold nanoparticles, silver nanoparticles, zinc oxide nanoparticles, and carbon nanotubes [18-49 26]. For our purpose, a nanoparticle (NP) is a nano-object for which all three external dimensions are in 50 the nanoscale [27]. Importantly, spICP-MS is sufficiently sensitive to measure nano-objects, including 51 NPs, at environmentally relevant concentrations (parts per trillion mass concentrations). Several factors 52 (e.g., signal dwell time, background correction algorithm, split-particle events, and nebulization 53 transport efficiency) are known to influence the accuracy of spICP-MS measurements [28]. While some 54 strategies to address such factors have been formulated, many improvements remain to be made [29-55 32]. For example, the split-particle correction approach devised and used in this laboratory has been 56 described in detail [30].

57 To our knowledge, the impact of drift in the sensitivity of the ICP-MS instrument on spICP-MS 58 analysis has not yet been studied. Drift can be defined as a continuous or incremental change in 59 response of a measuring instrument due to changes in the metrological properties of that measuring 60 instrument [33]. Drift in sensitivity is known to affect the accuracy of ICP-MS analysis generally and can 61 in some cases be quite severe. For example, a previous study from our laboratory showed a 62 spontaneous (i.e., not intentionally introduced) sensitivity drift for the isotope of interest of up to 50 % 63 during a 16 h period while employing a properly functioning instrument [34]. While most ICP-MS 64 analyses do not require such long operation and sensitivity drift is not often so severe, this observation 65 nonetheless points out that drift can be potentially problematic. Thus, the impact that sensitivity drift 66 could have on spICP-MS measurements could be substantial, but is until now unknown. One option to 67 mitigate the effect of drift during ICP-MS analysis is to include drift correction standards, either a 68 calibration standards or other standards, in the sample run order. However, this approach increases the 69 overall analysis time and does not enable a correction for each individual sample based on the drift at 70 the precise time that sample is being analyzed. Thus, correction for drift in ICP-MS analysis is often 71 performed by the use of an internal standard (ISD). Inclusion of a proper ISD can also help correct for 72 matrix effects. A recent study by Telgmann et al. [23] has used isotope dilution analysis to measure the 73 size distribution of AgNPs spiked with an enriched <sup>109</sup>Ag standard. This approach was effective for the 74 correction of matrix effects when testing the AgNPs in complex media such as wastewater and a river 75 water sample. However, the isotope dilution approach is only suitable for NPs composed of a principal 76 element having more than one isotope and for which there exists a readily available isotopically

77 enriched standard. It is not applicable to AuNPs, because Au is monoisotopic.

78 The aim of this paper is to investigate the potential impact of instrument sensitivity drift on the 79 accuracy of spICP-MS measurements of NP size and size distribution. This investigation was performed 80 both through theoretical modeling and laboratory experiments using National Institute of Standards and 81 Technology (NIST) reference material (RM) gold nanoparticles (AuNPs). To experimentally measure the 82 effect of drift on results in a controlled and quantifiable way, the instrument sensitivity was intentionally 83 decreased after instrument calibration, but prior to spICP-MS analysis of the RM AuNPs. To correct for 84 the significant observed impact of instrument drift, we investigated incorporation of two different ISDs. 85 To our knowledge, this is the first study to correct signal drift induced size bias by incorporation of an 86 ISD.

87

### 88 2 Materials and methods

# 2.1 Theoretical Modeling of Bias in Measured Diameter of Spherical NPs as a Function of Drift in ICP-MS Instrument Sensitivity

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Consider a spICP-MS measurement of NP diameter in a hypothetical suspension of ideal,
 monodisperse, non-agglomerated, non-aggregated, fully dense, spherical NPs. When the NPs are
 measured just after instrument calibration and with no change in the ICP-MS instrument sensitivity, let
 the measured NP mass be *m*<sub>1</sub>, where *d*<sub>1</sub> is the NP diameter, and *ρ* is the density:

96 
$$m_1 = \pi \frac{d_1^3}{6} \rho$$
 (1)

97

The technique of spICP-MS actually measures the mass of the nanoparticle, from which the diameter iscalculated by rearrangement of Eq. 1:

101 
$$d_1 = \sqrt[3]{\frac{6m_1}{\pi\rho}}$$
 (2)

103 Now, assume that the same suspension of NPs is measured again after the instrument sensitivity has 104 drifted by a percentage x (e.g., x = -20 % means that the instrument sensitivity has decreased by 20 %), 105 resulting in a biased NP mass  $m_2$ :

106

107 
$$m_2 = (1 + x/100)m_1$$
 (3)

108

#### 109 The diameter $d_2$ calculated from $m_2$ is then:

110

111 
$$d_2 = \sqrt[3]{\frac{6m_2}{\pi\rho}} = \sqrt[3]{\frac{6(1+x/100)m_1}{\pi\rho}}$$
 (4)

112

113 Therefore, the percentage *y* by which the diameter has been biased by the sensitivity drift is:

114

115 
$$y = 100 \left(\frac{d_2}{d_1} - 1\right) = 100 \left(\sqrt[3]{1 + \frac{x}{100}} - 1\right)$$
 (5)

116

For the example of x = -20 %, y = -7 %, meaning that the observed NP diameter theoretically will be biased low by 7 % in the presence of a 20 % reduction in ICP-MS instrument sensitivity that occurs after instrument calibration.

120 Theoretical modeling of this sort for several other spICP-MS measurements of nano-objects such 121 as cubes, rods, and plates is included in the Electronic Supplementary Material. From those theoretical 122 examples, it is shown that the magnitude of the bias in the measured nano-object dimension(s) induced 123 by instrument sensitivity drift depends on the number of dimensions being measured. Fig S1 in the 124 Electronic Supplementary Material illustrates that the most severe bias occurs for nano-objects having 125 one dimension in the nanoscale (or more precisely, when one dimension of a nano-object is being 126 measured). This is noteworthy, because the equivalent spherical diameter of nanoparticles, which is by 127 far the most common measurement made using spICP-MS, in essence involves all three dimensions.

128

#### 129 **2.2** Chemicals

Reagent grade high-purity deionized water (minimum resistivity of 18 MΩ cm) obtained from a
 ModuLab high-flow water purification system (Continental Water Systems, San Antonio, TX, USA) was

- used for all sample preparations and dilutions. Concentrated nitric acid (69 % *m/m*) and hydrochloric
- acid (32 % to 38 % *m/m*) (Veritas<sup>™</sup> double distilled, GFS Chemicals, Columbus, OH, USA) were used in
- 134 ICP-MS experiments. Gold nanoparticle reference materials with nominal diameters of 30 nm and 60 nm
- obtained from NIST (RM 8012 and RM 8013, respectively, Gaithersburg, MD, USA) were used in this
- 136 study. NIST Standard Reference Material<sup>®</sup> (SRM) 3121 Gold (Au) Standard Solution, SRM 3140 Platinum
- 137 (Pt) Standard Solution, and SRM 3124a Indium (In) Standard Solution were used to prepare solutions.
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# 139 2.3 Impact of Instrument Sensitivity Drift on spICP-MS Size Measurements of NIST 140 RM AuNPs

141 A Thermo XSERIES 7 quadrupole mass spectrometer (Waltham, MA, USA) equipped with a 142 concentric quartz C-Type nebulizer and Ni cones was used throughout this study for spICP-MS 143 measurements. Before analysis, the ICP-MS was tuned using a multi-element standard solution (2  $\mu$ g L<sup>-1</sup> 144 of each of Li, Be, Co, In, Ba, Ce and U in 2 % *v*/*v* HNO<sub>3</sub>) for maximum <sup>115</sup>In sensitivity and minimum oxide 145 (<sup>156</sup>CeO/<sup>140</sup>Ce) level (< 2 %). Data were collected at *m*/*z* 197 for Au. The sample flow rate was set to 0.6

146 mL min<sup>-1</sup> and measured daily in triplicate by weighing the water uptake after 5 min of aspiration.

147 All samples and working standard solutions were prepared gravimetrically (i.e., on a mass fraction 148 basis). AuNPs were suspended in a solution of deionized water at a particle number concentration between 2.5 x 10<sup>5</sup> g<sup>-1</sup> and 3.5 x 10<sup>5</sup> g<sup>-1</sup>. The AuNP stock suspensions were bath sonicated (model 2800, 149 Branson Ultrasonics, Danbury, CT, USA) for 10 min before dilution. Dissolved gold calibration standards 150 151 were prepared in a range of mass fraction between 0.05  $\mu$ g L<sup>-1</sup> and 5  $\mu$ g L<sup>-1</sup> either in an aqua 152 regia/thiourea solution (2.0 % *m/m* HCl, 0.5 % *m/m* HNO<sub>3</sub> and 0.5 % *m/m* thiourea) or in deionized water. To test the system repeatability, it was necessary to make several runs of the same sample, and 153 154 the uptake time was increased (60 s to 120 s) in order to eliminate memory effects and to properly 155 condition the introduction system. Additionally, thoroughly washing the system with 2 % v/v HNO<sub>3</sub> or

aqua regia/thiourea solution was necessary between each sample analysis.

157 To assess the impact of the instrument sensitivity drift on the accuracy of spICP-MS 158 measurements, the sensitivity of the ICP-MS was intentionally decreased after instrument calibration. 159 Instrument sensitivity was lowered by decreasing the absolute value of the extraction voltage from -600 160 V to -329 V for a 50 % decrease in the signal intensity. Results were also obtained for a 20 % decrease in the signal intensity through decreasing the detector voltage. The NIST RM AuNPs with nominal sizes of 161 30 nm and 60 nm were individually analyzed, first without sensitivity reduction and then with the 162 163 instrument sensitivity decreased by 20 % or 50 %. The time-resolved analysis (TRA) mode of the ICP-MS instrument was used for spICP-MS measurements. Raw data were recorded using Thermo Fisher 164 PlasmaLab software in units of counts per second (cps) and exported into Microsoft Excel for further 165 166 data processing. AuNP size calculations are described in depth in a later section.

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### 168 **2.4** Use of ISDs to Correct for ICP-MS Instrument Sensitivity Drift

169Two elements (In and Pt) were tested as ISDs to correct for the effect of changes in instrument170sensitivity on spICP-MS NP size measurements. Indium was tested, because it is widely used as an ISD

171 for routine ICP-MS analysis. Platinum was also tested, because it has mass-to-charge ratio (*m*/*z* 195) and

first ionization energy (8.9587 eV) values close to those of Au (*m*/*z* 197 and 9.2255 eV, respectively).

Data were collected at m/z 197 and m/z 195 for solutions with Au and Pt, or at m/z 197 and m/z 115 for

174 solutions with Au and In.

175 Because AuNPs were suspended in deionized water, ISDs were also prepared in water and needed 176 to be made freshly before addition to the samples, owing to poor long-term stability of the ISDs in 177 water. ISDs were either directly added to the sample during sample preparation (after dilution) or added 178 on-line using a second peristaltic pump channel and a tee placed directly before sample injection into 179 the nebulizer. The purpose of the tee placed before the nebulizer was to reduce the number of sample 180 preparation steps and to add the ISD immediately before injection, thus decreasing the interaction of 181 the ISD with the AuNPs. When adding the ISDs using the tee, an additional peristaltic pump was used to 182 pump out the waste from the spray chamber. When the ISDs were instead added directly to the sample 183 suspensions, the ISD stocks were first diluted in deionized water and then mixed with 30 nm and 60 nm 184 AuNP samples (diluted in water only) or with dissolved Au solution (diluted in water or in aqua 185 regia/thiourea) to a final ISD concentration of approximately 2  $\mu$ g L<sup>-1</sup>.

186 To assess the impact of the mixing methods on dissolved Au and AuNP measurements, standard 187 solutions of dissolved Au and suspensions of AuNPs were analyzed with introduction of the Pt ISD using 188 the tee or by direct mixing. For these experiments, the dissolved Au standards (0.05  $\mu$ g L<sup>-1</sup>, 0.5  $\mu$ g L<sup>-1</sup> and 1  $\mu$ g L<sup>-1</sup>) were analyzed by ICP-MS using continuous (i.e., not TRA) mode.

The measurement time for spICP-MS analysis was either 100 s or 400 s if one or two isotopes were analyzed, respectively. For most experiments involving the measurement of two isotopes, each dwell time was set to 10 ms, with the quadrupole settling time set to 10 ms between each peak hop. Thus, it was necessary to increase the sample analysis time by a factor of 4 compared to analyzing the AuNPs alone, to allow for the same number of measurement events when analyzing both the ISD and the AuNPs.

196

### 197 2.5 Calculation of AuNP Size

198 The theoretical equations used in this study are derived from those presented by Pace *et al.*[31] 199 The nanoparticle signal spikes were identified as those with intensities exceeding five times the standard 200 deviation of the background ( $5\sigma$ ) [32]. The determination of the mass of the Au nanoparticle  $m_{Au}$ 201 causing a signal spike is described by Eq. 6:

202

203

$$m_{Au} = \frac{S_{Au}}{slope} \tag{6}$$

204

where S<sub>Au</sub> is the background-subtracted intensity of a signal spike and *slope* is the slope of the
calibration line generated using dissolved Au solutions. For this purpose, the calibration line is defined
using intensity as the ordinate and the mass of Au entering the plasma within a dwell time as the
abscissa. The latter, *m*<sub>Au,dwell</sub>, is given by:

 $m_{Au,dwell} = C_{Au} * t_{dwell} * q_{liq} * \eta_n$ 

211

212 where  $C_{Au}$  is the mass concentration of Au in the given solution being nebulized,  $t_{dwell}$  is the dwell time,  $q_{lig}$  is the solution nebulization rate, and  $\eta_n$  is the nebulization transport efficiency, which is the fraction 213 214 of nebulized Au that actually enters the plasma. The nebulization transport efficiency was determined 215 each day by using the particle size method of Pace et al.[31] This approach relies on well-characterized reference nanoparticles of a known particle size. Typically,  $\eta_n$  was about 2 %. The use of a tee piece can 216 change the flow rate introduced in the nebulizer and consequently the nebulization efficiency. 217 218 Nevertheless, the calculations of the size distribution of NPs remain unchanged. Additionally, it has been 219 observed that the sensitivity decrease did not impact the nebulization efficiency. For instance,  $\eta_n$ 220 calculated from the particle frequency was  $(0.9 \pm 0.1)$  % and  $(1.0 \pm 0.1)$  % before and after 50 % 221 decrease, respectively.

222 When using ISD correction, the background-corrected intensity of the ISD  $S_{ISD}$  and the mass of 223 the ISD that enters the plasma in a single dwell time  $m_{ISD,dwell}$  must be taken into account:

224

225 
$$m_{Au} = \frac{S_{Au}/S_{ISD}}{slope} m_{ISD,dwell}$$

226

where the slope is determined from the calibration line of dissolved Au ( $S_{Au}/S_{ISD}$  as a function of  $m_{Au}/m_{ISD}$ ). The parameter  $m_{ISD,dwell}$  is defined analogously to Eq. 7.

229 For the data obtained in our study, the ratio of  $S_{Au}$  to  $S_{ISD}$  cannot be calculated for each individual dwell time window. This is because  $S_{Au}$  and  $S_{ISD}$  were measured sequentially, not simultaneously, as a 230 231 result of the instrument peak hopping back and forth between the isotopes. The data stream in TRA 232 mode of  $S_{Au}$  and  $S_{ISD}$  values was divided into segments for data processing. Each segment contained 100 233 dwell time windows for S<sub>Au</sub> and 100 dwell time windows for S<sub>ISD</sub>. For each segment, S<sub>ISD</sub> was averaged over the 100 dwell times, and each  $S_{Au}$  value in the segment was divided by this average  $S_{ISD}$  value. As an 234 example, a dwell time of 10 ms for both <sup>197</sup>Au and <sup>195</sup>Pt and a total measurement time of 400 s resulted 235 236 in 200 segments of 2 s each, with each segment containing 100 dwell time windows of the  $S_{Au}$  signal. The 237 segment length (100 dwell times) was chosen judiciously to provide some signal averaging (preventing 238 too much noise from the signal fluctuation intrinsically related to the instrument), while avoiding serious 239 depletion of the effectiveness of the ISD to correct for instrument sensitivity drift.

The mean of the size distribution for each run of a given sample suspension was determined as the mean of a Gaussian curve fitted to the histogram using OriginPro 9.1. The uncertainty values given throughout the manuscript are the standard deviations of the Gaussian fitting provided by the OriginPro processing (Gaussian fitting uncertainty). Additionally, estimations of measurement repeatability were determined from at least two replications of each suspension. In this case, the uncertainty was calculated as the standard deviation of the mean of the replicate values.

(7)

(8)

### 247 3 Results and discussion

## 3.1 Impact of decreased instrument sensitivity on the NP size from theoretical modeling

250 The dependence of the percentage change in equivalent spherical diameter on the percentage of 251 instrument sensitivity drift is plotted in Fig. 1. Decreases in instrument sensitivity of 20 % and 50 % result 252 in NP diameters that are 7.2 % and 21 % smaller than the actual size, respectively. While it may seem at 253 first glance that the particle size should decrease by 20 % when the instrument sensitivity decreases by 254 that amount, what decreases by 20 % is the measured intensity, which is proportional to the NP mass 255 and to the cube root of the NP diameter. Nevertheless, the impact of instrument sensitivity change on 256 spICP-MS measurements is predicted to substantially impact the results obtained. Sensitivity gains are 257 far less frequent than losses, but are included in the figure for completeness. The case of measuring 258 equivalent spherical diameter depicted in Fig. 1 involves all three external dimensions, because 259 diameter necessarily defines all three dimensions. Theoretical modeling results for several other types 260 of spICP-MS dimensional measurements of nano-objects, along with plots similar to the one shown in 261 Fig. 1 for measurements of two dimensions and one dimension, are given in the Electronic 262 Supplementary Material (see particularly Fig. S1).

263 Changes of instrument sensitivity of 20 % and 50 % are severe, although they have been observed 264 to occur spontaneously in this laboratory [34]. If the sensitivity drifted by 10 % to 15 %, an amount 265 somewhat regularly observed in our laboratory conducting ICP-MS analysis during an 8 h to 10 h period, 266 this would result in a spICP-MS size bias of approximately 3 % to 5 % for spheres (Fig. 1) and larger 267 biases for other shapes (Electronic Supplementary Material Fig. S1). While other sources of variability 268 and bias in spICP-MS may currently be as or more significant than the bias from instrument drift, 269 correcting for this bias will become increasingly important as the technique matures and other 270 uncertainty sources are accounted for and reduced.

271

### 272 **3.2** Impact of instrument sensitivity drift on measured AuNP sizes

273 The impact of decreased instrument sensitivity, as has been documented to occur during 274 operation of ICP-MS instruments [34], on the calculated AuNP sizes was also experimentally evaluated. 275 The measured equivalent spherical diameters of the NIST 30 nm and 60 nm AuNPs without a change in 276 the instrument sensitivity were  $(27.6 \pm 1.7)$  nm and  $(56.2 \pm 2.8)$  nm, respectively (Fig. 2). These 277 uncertainty values account only for the standard deviation of the Gaussian fitting. The average sizes 278 determined by transmission electron microscopy (TEM) and provided in the RM reports of investigation 279 are  $(27.6 \pm 2.1)$  nm and  $(56.0 \pm 0.5)$  nm for the nominal 30 nm and 60 nm diameter AuNPs, respectively. 280 The uncertainties for these TEM results are expanded to a level of confidence of approximately 95 % 281 [35,36]. The spICP-MS results are in good agreement with the TEM results.

282The impact of sensitivity decreases of 20 % and 50 % on the measured sizes was also evaluated283(Fig. 2). For a 20 % decrease in sensitivity, the equivalent spherical diameters of the 30 nm and 60 nm284AuNPs were (25.3 ± 1.7) nm and (51.7 ± 2.3) nm, respectively, equivalent to percentage biases of -7.6 %

and -8.4 %, respectively. There was a larger bias in the measured diameters when the sensitivity was decreased by 50 %. Using this condition, the measured sizes were  $(20.0 \pm 2.3)$  nm and  $(42.8 \pm 3.4)$  nm for

the 30 nm and 60 nm AuNPs, respectively, equivalent to an approximate -25 % bias for both AuNPs.
These observed biases are quite close to the theoretically predicted values of -7 % and -21 % for

decreases in instrument sensitivity of 20 % and 50 %, respectively, calculated earlier in this paper.

290 Overall, these data suggest that changes in instrument sensitivity after calibration can 291 substantially impact measured nanoparticle sizes, and that not accounting for changes in sensitivity can 292 lead to biased results. In the subsequent sections, we describe an investigation of the use of ISDs to

293 correct for changes in instrument sensitivity on spICP-MS dimensional measurements.

294

### 295 3.3 Selection and stability of the ISD

When employing an ISD in spICP-MS, it is appropriate for the ISD to be in a stock solution that has a similar matrix to the solution phase of the NP suspensions to be analyzed, because NPs may be unstable relative to changes in solution matrix. The ISD must also be stable in solution both with and without the presence of NPs. For both of these factors, stability should be much more important when the sample suspension and ISD are directly mixed than when mixing via the tee is employed, owing to the much longer contact time between the sample and ISD. Pt and In were each evaluated for use as an ISD applied to spherical diameter measurements of AuNPs.

303 Indium quickly proved to be unsuitable for this particular application. While this metal is known to 304 be unstable at neutral pH, it was tested nonetheless, because of the possibility that the kinetics of 305 instability would be slow enough to permit its use. However, signal spikes were observed for In in water 306 (Electronic Supplementary Material Fig. S2a), whether it was mixed with AuNPs or not, and for both 307 direct mixing and online ISD introduction using the tee. The signal spikes may be due to formation of 308 indium oxide particles and precipitates [37,38]. Given these findings, use of In as an ISD was abandoned. 309 In contrast to In, Pt provided stable signals (RSD approximately 6 %, Electronic Supplementary Material 310 Fig S2b) in the presence and absence of AuNPs, using both direct mixing and online introduction. 311 Possible explanations for the Pt stability are the formation of hydroxylated species of Pt at neutral pH 312 [39] or sufficiently slow instability kinetics. Because of its superior stability, Pt was utilized as the ISD for 313 the remainder of this study.

314

# 315 3.4 Influence of ICP-MS parameters on AuNP size measurements with Pt internal 316 standardization

The impact of the Pt ISD on the AuNP size measurements in the absence of instrument sensitivity drift was analyzed using different ICP-MS parameters. Results for testing nominal 30 nm and 60 nm NIST RM AuNPs while measuring only Au for an analysis period of 100 s or Au and Pt for a total analysis time of 400 s are presented in Figs. S3a and S3b of the Electronic Supplementary Material. For these measurements, a dwell time of 10 ms for both Au and Pt and a quadrupole settling time of 10 ms were used. The frequencies of AuNP events were equivalent with and without Pt monitoring for both the 30 nm and 60 nm AuNPs. The mean equivalent spherical diameter values for the 30 nm and 60 nm AuNP distributions were (27.6  $\pm$  1.7) nm and (56.2  $\pm$  2.8) nm, respectively, when an analysis time of 100 s was used and only Au was analyzed. These values are very similar to those measured during a measurement time of 400 s with simultaneous analysis of Pt and Au, (27.2  $\pm$  1.9) nm and (56.1  $\pm$  2.9) nm, respectively. Thus, no significant change in measured size was observed when these two elements were analyzed simultaneously.

329 The impact of increasing the analysis speed by decreasing the quadrupole settling time from 10 330 ms to 5 ms or decreasing the dwell time for Pt from 10 ms to 1 ms on the spICP-MS measurement was 331 also assessed. While the results showed that the average signal (in cps) for a dissolved solution of Pt is 332 similar for both dwell times, the RSD of the Pt signal was twice as high for a dwell time of 1 ms (RSD = 12 333 %) compared to that for a dwell time of 10 ms (RSD = 5.6 %). Thus, in this particular case using a higher 334 dwell time for the ISD improved measurement precision. This finding is expected because the number of 335 ions counted with a dwell time of 1 ms is ten times lower than at 10 ms, and Poisson noise is equal to 336 the square root of the number of ions counted [40]. However, the size of AuNPs determined by this 337 method was not modified given that the SISD averages were similar; it should be noted that the dwell 338 time for the AuNPs always remained at 10 ms. It might be possible to use shorter dwell times for the ISD 339 without the increased measurement variability by increasing the Pt concentration. However, it is also 340 possible that the increased interactions between Pt and the AuNPs could lead to deleterious effects.

A decrease of the quadrupole settling time from 10 ms to 5 ms did not significantly change the size distribution (Electronic Supplementary Material Fig. S4). The mean sizes for the 30 nm standard were (27.6 ± 2.2) nm and (27.8 ± 2.0) nm for 5 ms and 10 ms settling times, respectively, while the mean sizes for the 60 nm standard were (57.2 ± 3.2) nm and (57.5 ± 3.4) nm, respectively. The small *m/z* difference between the Au and Pt isotopes apparently enabled the quadrupole to stabilize adequately with the shorter settling time.

347 The impact of introduction of the ISD using direct mixing or the tee on the analysis of the 348 dissolved gold and AuNPs was also investigated. The dissolved Au signal decreased by a factor of  $1.9 \pm$ 0.1 when the tee was used as compared to the direct mixing approach. This result can be explained by 349 350 the dilution of the dissolved gold concentration when mixing equivalent volumes of the dissolved gold 351 solution and the ISD solution when using the tee. Indeed, due to a change in the flow rate the 352 nebulization efficiency was modified (from approximately 2 % to about 1 %). Nevertheless, the mean 353 intensity of AuNP spikes (about 8000 cps for 30 nm and 60000 cps for 60 nm AuNPs), and consequently 354 the measured AuNP size, are similar for both ISD introduction approaches as expected.

# 355 3.5 Correction of instrument sensitivity drift in AuNP size measurement with use of an ISD

The effectiveness of Pt as an ISD to correct for changes in instrument sensitivity was evaluated. It was 357 358 important to first assess the impact of ISD correction in the absence of instrument sensitivity drift (i.e., 359 immediately after calibration and without any observable drift). When operating the ICP-MS under this 360 condition, the average size obtained for the 30 nm AuNPs was  $(27.2 \pm 2.0)$  nm and  $(27.9 \pm 1.9)$  nm 361 without and with ISD correction, respectively; results obtained for the 30 nm and 60 nm AuNPs without 362 ISD correction are the same as those described in section 3.4 with a measurement time of 400 s and are 363 repeated here to enable a direct comparison to the results with the ISD correction. For the 60 nm AuNPs 364 the average particle size was  $(56.1 \pm 2.9)$  nm and  $(55.9 \pm 3.1)$  nm without and with ISD correction,

respectively (see Table 1 and Electronic Supplementary Material Figs. S3b and S3c). Therefore, it is
 reasonable to conclude that no significantly deleterious effects were observed for the use of Pt as the
 ISD and the ISD correction.

368 The size distributions of 30 nm and 60 nm AuNPs after an induced transient 20 % loss of 369 sensitivity were analyzed with and without ISD correction. As shown in Table 1 and described earlier, the 370 mean sizes measured by spICP-MS were underestimated when the ISD was not used, and the relative 371 magnitudes of the underestimations are in agreement with theory. However, when Pt was employed as 372 the ISD, the mean sizes were corrected and in agreement with the expected TEM mean sizes. A more 373 extreme loss of sensitivity of 50 % was also tested by decreasing the extraction voltage; these 374 experiments were conducted by using the tee piece, a modification shown in section 3.4 to not impact 375 spICP-MS results. While a 50 % decrease in sensitivity may seem like an excessive change, instrument 376 drift of this magnitude was observed in a previous study [34]. As shown in Table 1, the results after ISD 377 correction are very similar to those obtained for the 30 nm AuNPs before the instrument sensitivity 378 decrease was induced. The decreased instrument sensitivity caused the mean sizes of the NPs to be 379 underestimated to magnitudes that agree with theoretical modelling results, and use of the ISD 380 effectively corrected the bias. The size distributions for the spICP-MS results for the 30 nm and 60 nm 381 AuNPs on which Table 1 is based are presented in Figs. 3 and 4, respectively.

382 Some of the newest quadrupole ICP-MS instruments allow one to utilize very short dwell times on 383 the order of microseconds, with quadrupole settling times that can also be very short or even omitted. 384 With such capabilities, the method developed in this study could be applied using an almost 385 simultaneous ratio of analyte and internal standard signal intensities during the analysis of individual 386 nanoparticles, producing results that might be more accurate. As a way to simulate crudely this type of 387 capability, the results in Fig. 3 were recalculated without averaging the Pt signal over 100 dwell time 388 windows, as described in section 2.5, but by pairing each individual Pt intensity measurement with its 389 adjoining Au intensity measurement (Electronic Supplementary Material Fig. S5). When visually 390 compared, the results computed in this way do not seem to be better than those in Fig. 3. As noted, 391 however, this is a crude simulation at best, and actual tests on an instrument with the advanced 392 capabilities will need to be performed at a later date. Interestingly, an instrument capable of conducting 393 rapid peak hopping opens the possibility of performing multielement analyses on single nanoparticles, 394 which was impossible with quadrupoles only a few years ago.

395 Of course, essentially truly simultaneous measurements of multiple isotopes are possible with 396 ICP-MS technologies that do not employ quadrupoles, and the ISD approach demonstrated here could 397 certainly be applied. However, each alternative has disadvantages. Time-of-flight (TOF) instruments 398 have been utilized for simultaneous, multielement analyses for many years, but these instruments 399 generally have relatively poor sensitivity and limits of detection (LODs). Multicollector instruments offer 400 very good sensitivity and LODs, but are extremely expensive and are constrained in the range of mass-401 to-charge ratios that can be monitored in a single experiment, thereby limiting the possible choices of 402 internal standard.

It would, in theory, be possible to correct for instrument drift using enriched isotope standards of
the same element, an approach that was previously used to correct matrix effects in analyses of AgNPs
[23]. However, this approach is impossible when the analyte is monoisotopic or when a suitable
isotopically enriched standard is unavailable.

For the study reported in this paper, AuNPs were selected as a suitable test case. The approach described for drift correction obviously should be applicable to spICP-MS analyses of NPs other than AuNPs. In any case it is important to conduct preliminary experiments as described above for Pt with AuNPs to assess the necessary quadrupole settling time, variability in the ISD signal, and stability of the ISD in the matrix.

### 412 4 Conclusions

413 The study described in this paper shows clearly that a change in ICP-MS instrument sensitivity 414 after calibration can induce bias in the measured NP size distributions and in the mean sizes derived 415 from those distributions. It also demonstrates effective correction of this bias through the use of a well-416 chosen ISD. The size distribution calculation relies on a calibration curve obtained after analysis of 417 dissolved Au standards [31] with incorporation of the ISD and before any significant change in 418 sensitivity. Therefore, while the average AuNP size measured without the ISD was biased by the change 419 in the instrument sensitivity, the concomitant change in the ISD signal intensity could be used to correct 420 for the instrument sensitivity loss. The ISD can also help to correct for matrix effects, which are of 421 particular interest when analyzing complex samples (environmental or biological) and when calibrating 422 using calibrants that have a matrix that is not precisely the same as the matrix of the unknown sample. 423 One limitation of the use of an ISD is that split-pulse correction, as devised and implemented in this 424 laboratory [30], is not possible, because two consecutive pulses in the data set do not correspond to two 425 consecutive time periods in the analysis. Recent innovations in spICP-MS equipment (e.g., quadrupole 426 instruments with very short dwell time and rapid peak hopping capabilities [41] and ICP-TOF-MS [42]) 427 are much less prone to measurement bias induced by split particle events.

428

#### 429 Electronic Supplementary Material Available

430 Theoretical modeling of the impact of instrument sensitivity drift on sp-ICP-MS measurements of the

dimension(s) of nano-objects, and figures showing the theoretical bias caused by instrument drift for

432 nano-objects with different numbers of dimensions, ICP-MS intensity of indium in an aqueous solution

433 without nanoparticles, impact of different analysis times with and without using Pt as an internal

- 434 standard for correction on AuNP size distributions, and the impact of quadrupole settling time on AuNP
- 435 size distributions.
- 436

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- 440 necessarily the best available for the purpose.
- 441

#### 442 Compliance With Ethical Standards

443 **Conflict of interest** The authors declare that they have no conflict of interest.

### 445 **References**

- 1. ISO/TS 80004-1:2010 Nanotechnologies Vocabulary Part 1: Core Terms (2010). International
  Organization for Standardization, Geneva, Switzerland
- 448 2. Lee J, Mahendra S, Alvarez PJJ (2010) Nanomaterials in the Construction Industry: A Review of Their
- Applications and Environmental Health and Safety Considerations. ACS Nano 4 (7):3580-3590.
  doi:10.1021/nn100866w
- 451 3. Bauer F, Flyunt R, Czihal K, Ernst H, Naumov S, Buchmeiser MR (2007) UV curing of nanoparticle
- reinforced acrylates. Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions
   with Materials and Atoms 265 (1):87-91. doi:http://dx.doi.org/10.1016/j.nimb.2007.08.030
- 454 4. Kamat PV (2012) Boosting the Efficiency of Quantum Dot Sensitized Solar Cells through Modulation of 455 Interfacial Charge Transfer. Accounts of Chemical Research 45 (11):1906-1915. doi:10.1021/ar200315d
- 456 5. Petersen EJ, Pinto RA, Shi X, Huang Q (2012) Impact of size and sorption on degradation of
- 457 trichloroethylene and polychlorinated biphenyls by nano-scale zerovalent iron. Journal of Hazardous
- 458 Materials 243 (0):73-79. doi:<u>http://dx.doi.org/10.1016/j.jhazmat.2012.09.070</u>
- 459 6. Klaine SJ, Koelmans AA, Horne N, Carley S, Handy RD, Kapustka L, Nowack B, von der Kammer F (2012)
  460 Paradigms to assess the environmental impact of manufactured nanomaterials. Environmental
  461 Tavian and Chamistry 21 (1):2 14 dai:10.1002 (sta 722)
- 461 Toxicology and Chemistry 31 (1):3-14. doi:10.1002/etc.733
- 7. Nowack B, Ranville JF, Diamond S, Gallego-Urrea JA, Metcalfe C, Rose J, Horne N, Koelmans AA, Klaine
   SJ (2012) Potential scenarios for nanomaterial release and subsequent alteration in the environment.
- 464 Environmental Toxicology and Chemistry 31 (1):50-59. doi:10.1002/etc.726
- 465 8. Oomen AG, Bos PMJ, Fernandes TF, Hund-Rinke K, Boraschi D, Byrne HJ, Aschberger K, Gottardo S,
- 466 von der Kammer F, Kühnel D, Hristozov D, Marcomini A, Migliore L, Scott-Fordsmand J, Wick P,
- 467 Landsiedel R (2014) Concern-driven integrated approaches to nanomaterial testing and assessment –
- 468 report of the NanoSafety Cluster Working Group 10. Nanotoxicology 8 (3):334-348.
- 469 doi:doi:10.3109/17435390.2013.802387
- 470 9. Petersen EJ, Zhang L, Mattison NT, O'Carroll DM, Whelton AJ, Uddin N, Nguyen T, Huang Q, Henry TB,
- 471 Holbrook RD, Chen KL (2011) Potential Release Pathways, Environmental Fate, And Ecological Risks of
- 472 Carbon Nanotubes. Environmental Science & Technology 45 (23):9837-9856. doi:10.1021/es201579y
- 473 10. Krug HF, Wick P (2011) Nanotoxicology: An Interdisciplinary Challenge. Angewandte Chemie
  474 International Edition 50 (6):1260-1278. doi:10.1002/anie.201001037
- 475 11. Hassellöv M, Readman J, Ranville J, Tiede K (2008) Nanoparticle analysis and characterization
- 476 methodologies in environmental risk assessment of engineered nanoparticles. Ecotoxicology 17 (5):344477 361. doi:10.1007/s10646-008-0225-x
- 478 12. Gallego-Urrea JA, Tuoriniemi J, Hassellöv M (2011) Applications of particle-tracking analysis to the
- 479 determination of size distributions and concentrations of nanoparticles in environmental, biological and
- 480 food samples. TrAC Trends in Analytical Chemistry 30 (3):473-483.
- 481 doi:<u>http://dx.doi.org/10.1016/j.trac.2011.01.005</u>

- 482 13. Pace HE, Rogers NJ, Jarolimek C, Coleman VA, Gray EP, Higgins CP, Ranville JF (2012) Single Particle
- 483 Inductively Coupled Plasma-Mass Spectrometry: A Performance Evaluation and Method Comparison in
- the Determination of Nanoparticle Size. Environmental Science & Technology 46 (22):12272-12280.
  doi:10.1021/es301787d
- 486 14. Petersen EJ, Henry TB, Zhao J, MacCuspie RI, Kirschling TL, Dobrovolskaia MA, Hackley V, Xing B,
- 487 White JC (2014) Identification and Avoidance of Potential Artifacts and Misinterpretations in
- 488 Nanomaterial Ecotoxicity Measurements. Environmental Science & Technology 48 (8):4226-4246.
- 489 doi:10.1021/es4052999
- 490 15. Petersen EJ, Diamond SA, Kennedy AJ, Goss GG, Ho K, Lead J, Hanna SK, Hartmann NB, Hund-Rinke K,
- 491 Mader B, Manier N, Pandard P, Salinas ER, Sayre P (2015) Adapting OECD Aquatic Toxicity Tests for Use
- 492 with Manufactured Nanomaterials: Key Issues and Consensus Recommendations. Environmental Science
  493 & Technology 49 (16):9532-9547. doi:10.1021/acs.est.5b00997
- 494 16. Degueldre C, Favarger PY (2003) Colloid analysis by single particle inductively coupled plasma-mass
- 495 spectroscopy: a feasibility study. Colloids and Surfaces A: Physicochemical and Engineering Aspects 217 496 (1, 2):127,142, doi:http://dv.doi.org/10.1016/s0027.7757/02)00568, X
- 496 (1–3):137-142. doi:<u>http://dx.doi.org/10.1016/S0927-7757(02)00568-X</u>
- 497 17. Degueldre C, Favarger PY, Bitea C (2004) Zirconia colloid analysis by single particle inductively
- 498 coupled plasma–mass spectrometry. Analytica Chimica Acta 518 (1–2):137-142.
- 499 doi:<u>http://dx.doi.org/10.1016/j.aca.2004.04.015</u>
- 18. Reed RB, Goodwin DG, Marsh KL, Capracotta SS, Higgins CP, Fairbrother DH, Ranville JF (2013)
- Detection of single walled carbon nanotubes by monitoring embedded metals. Environmental Science:
   Processes & Impacts 15 (1):204-213. doi:10.1039/C2EM30717K
- 503 19. Mitrano DM, Lesher EK, Bednar A, Monserud J, Higgins CP, Ranville JF (2012) Detecting
- nanoparticulate silver using single-particle inductively coupled plasma-mass spectrometry.
   Environmental Toxicology and Chemistry 31 (1):115-121. doi:10.1002/etc.719
- 20. Degueldre C, Favarger PY, Wold S (2006) Gold colloid analysis by inductively coupled plasma-mass
   spectrometry in a single particle mode. Analytica Chimica Acta 555 (2):263-268.
- 508 doi:<u>http://dx.doi.org/10.1016/j.aca.2005.09.021</u>
- 509 21. Gray EP, Coleman JG, Bednar AJ, Kennedy AJ, Ranville JF, Higgins CP (2013) Extraction and Analysis of
- 510 Silver and Gold Nanoparticles from Biological Tissues Using Single Particle Inductively Coupled Plasma
- 511 Mass Spectrometry. Environmental Science & Technology 47 (24):14315-14323. doi:10.1021/es403558c
- 512 22. Laborda F, Bolea E, Jiménez-Lamana J (2013) Single Particle Inductively Coupled Plasma Mass
- 513 Spectrometry: A Powerful Tool for Nanoanalysis. Analytical Chemistry 86 (5):2270-2278.
- 514 doi:10.1021/ac402980q
- 515 23. Telgmann L, Metcalfe CD, Hintelmann H (2014) Rapid size characterization of silver nanoparticles by
  516 single particle ICP-MS and isotope dilution. Journal of Analytical Atomic Spectrometry 29 (7):1265-1272.
  517 doi:10.1039/C4JA00115J
- 518 24. Peters RB, Rivera Z, van Bemmel G, Marvin HP, Weigel S, Bouwmeester H (2014) Development and
- validation of single particle ICP-MS for sizing and quantitative determination of nano-silver in chicken
- 520 meat. Anal Bioanal Chem 406 (16):3875-3885. doi:10.1007/s00216-013-7571-0

- 521 25. Hineman A, Stephan C (2014) Effect of dwell time on single particle inductively coupled plasma mass
- 522 spectrometry data acquisition quality. Journal of Analytical Atomic Spectrometry 29 (7):1252-1257.
- 523 doi:10.1039/C4JA00097H
- 524 26. Linsinger TJ, Peters R, Weigel S (2014) International interlaboratory study for sizing and
- quantification of Ag nanoparticles in food simulants by single-particle ICPMS. Anal Bioanal Chem 406
   (16):3835-3843. doi:10.1007/s00216-013-7559-9
- 527 27. Liber K, Doig LE, White-Sobey SL (2011) Toxicity of uranium, molybdenum, nickel, and arsenic to
  528 Hyalella azteca and Chironomus dilutus in water-only and spiked-sediment toxicity tests. Ecotoxicol
  529 Environ Saf 74 (5):1171-1179. doi:10.1016/j.ecoenv.2011.02.014
- 530 28. Montoro Bustos AR, Petersen EJ, Possolo A, Winchester MR (2015) Post hoc Interlaboratory
- 531 Comparison of Single Particle ICP-MS Size Measurements of NIST Gold Nanoparticle Reference
- 532 Materials. Analytical Chemistry 87 (17):8809-8817. doi:10.1021/acs.analchem.5b01741
- 533 29. Laborda F, Jimenez-Lamana J, Bolea E, Castillo JR (2013) Critical considerations for the determination
- of nanoparticle number concentrations, size and number size distributions by single particle ICP-MS.
- 535 Journal of Analytical Atomic Spectrometry 28 (8):1220-1232. doi:10.1039/C3JA50100K
- 536 30. Liu J, Murphy KE, MacCuspie RI, Winchester MR (2014) Capabilities of Single Particle Inductively
- 537 Coupled Plasma Mass Spectrometry for the Size Measurement of Nanoparticles: A Case Study on Gold
- 538 Nanoparticles. Analytical Chemistry 86 (7):3405-3414. doi:10.1021/ac403775a
- 31. Pace HE, Rogers NJ, Jarolimek C, Coleman VA, Higgins CP, Ranville JF (2011) Determining Transport
   Efficiency for the Purpose of Counting and Sizing Nanoparticles via Single Particle Inductively Coupled
- 541Plasma Mass Spectrometry. Analytical Chemistry 83 (24):9361-9369. doi:10.1021/ac201952t
- 542 32. Tuoriniemi J, Cornelis G, Hassellöv M (2012) Size Discrimination and Detection Capabilities of Single543 Particle ICPMS for Environmental Analysis of Silver Nanoparticles. Analytical Chemistry 84 (9):3965544 3972. doi:10.1021/ac203005r
- 33. ISO (2008) International Vocabulary of Basic and General Terms in Metrology Basic and general
   concepts and associated terms (VIM), 3rd edition.108
- 34. Turk GC, Yu LL, Salit ML, Guthrie WF (2001) Using inductively coupled plasma–mass spectrometry for
  calibration transfer between environmental CRMs. Fresenius J Anal Chem 370 (2-3):259-263.
  doi:10.1007/s002160100790
- 35. NIST (2012) Reference Material<sup>®</sup> 8012 Gold Nanoparticles, Nominal 30 nm Diameter. National
   Institut of Standards and Technology,
- 36. NIST (2012) Reference Material<sup>®</sup> 8013 Gold Nanoparticles, Nominal 60 nm Diameter. National
   Institut of Standards and Technology,
- 37. Moeller T (1941) Contributions to the Chemistry of Indium. III. An Electrometric Study of the
  Precipitation of Hydrous Indium Hydroxide1. Journal of the American Chemical Society 63 (10):26252628. doi:10.1021/ja01855a026
- 38. Busev AI (2013) The Analytical Chemistry of Indium: International Series of Monographs onAnalytical Chemistry. Elsevier Science,

- 39. Azaroual M, Romand B, Freyssinet P, Disnar J-R (2001) Solubility of platinum in aqueous solutions at
- 560 25°C and pHs 4 to 10 under oxidizing conditions. Geochimica et Cosmochimica Acta 65 (24):4453-4466.
- 561 doi:<u>http://dx.doi.org/10.1016/S0016-7037(01)00752-9</u>
- 40. Laborda F, Medrano J, Castillo JR (2001) Quality of quantitative and semiquantitative results in
- inductively coupled plasma mass spectrometry. Journal of Analytical Atomic Spectrometry 16 (7):732 738. doi:10.1039/B101814K
- 565 41. Montano MD, Badiei HR, Bazargan S, Ranville JF (2014) Improvements in the detection and
- characterization of engineered nanoparticles using spICP-MS with microsecond dwell times.
   Environmental Science: Nano 1 (4):338-346. doi:10.1039/C4EN00058G
- 568 42. Borovinskaya O, Gschwind S, Hattendorf B, Tanner M, Günther D (2014) Simultaneous Mass
- 569 Quantification of Nanoparticles of Different Composition in a Mixture by Microdroplet Generator-
- 570 ICPTOFMS. Analytical Chemistry 86 (16):8142-8148. doi:10.1021/ac501150c
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573	Figure captions
574	
575	Fig. 1: Theoretical change in the equivalent spherical diameter of nanoparticles measured by
576	spICP-MS caused by drift in the sensitivity of the instrument
577	
578	Fig. 2: Size distributions of AuNPs with nominal diameters of a) 30 nm and b) 60 nm measured by
579	spICP-MS under normal operating conditions, with a 20 % loss of sensitivity, and with a 50 % loss
580	of sensitivity (at least two replicates were combined for each histogram)
581	
582	Fig. 3: Size distributions of AuNPs with nominal diameters of a) 30 nm and b) 60 nm in the absence of
583	instrument sensitivity drift and with and without ISD correction after a sensitivity decrease of 20 %.
584	Duplicate replicates were combined for each histogram. The measurement time was 400 s; dwell times
585	were 10 ms for both Au and Pt; quadrupple settling time was 10 ms. For the No decrease data the
587	sample solution before analysis.
588	

Fig. 4: Size distributions of AuNPs with nominal diameters of a) 30 nm and b) 60 nm in the absence of
 instrument sensitivity drift and with and without ISD correction after a sensitivity decrease of 50 %.
 Triplicate replicates were combined for each histogram. The measurement time was 200 s; dwell times

592 were 10 ms for Au and 1 ms for Pt; quadrupole settling time was 10 ms. For the "No decrease" data, the

samples were analyzed just before the loss of sensitivity was induced. Pt was added with the tee.





Fig. 1: Theoretical change in the equivalent spherical diameter of nanoparticles measured byspICP-MS caused by drift in the sensitivity of the instrument



Fig. 2: Size distributions of AuNPs with nominal diameters of a) 30 nm and b) 60 nm measured by spICP-MS under normal operating conditions, with a 20 % loss of sensitivity, and with a 50 % loss of sensitivity (at least two replicates were combined for each histogram) 





**Fig. 3**: Size distributions of AuNPs with nominal diameters of a) 30 nm and b) 60 nm in the absence of instrument sensitivity drift and with and without ISD correction after a sensitivity decrease of 20 %. Duplicate replicates were combined for each histogram. The measurement time was 400 s; dwell times were 10 ms for both Au and Pt; quadrupole settling time was 10 ms. For the "No decrease" data the samples were analyzed just before the loss of sensitivity was induced. Pt was added directly into the sample solution before analysis

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**Fig. 4**: Size distributions of AuNPs with nominal diameters of a) 30 nm and b) 60 nm in the absence of instrument sensitivity drift and with and without ISD correction after a sensitivity decrease of 50 %. Triplicate replicates were combined for each histogram. The measurement time was 200 s; dwell times were 10 ms for Au and 1 ms for Pt; quadrupole settling time was 10 ms. For the "No decrease" data, the samples were analyzed just before the loss of sensitivity was induced. Pt was added with the tee.

Table 1: Impact of ICP-MS instrument sensitivity drift on NP size measured with spICP-MS, and correction of the effect using Pt as an ISD

			Uncertainty of the Gaussian fitting		spICP-MS repeatability Mean Size (nm) <sup>a</sup>		Bias in Mean Size Without ISD	
AuNPs	TEM Mean Size (nm) <sup>b</sup>	Sensitivity Decrease <sup>c</sup>	With Pt ISD	Without Pt ISD	With Pt ISD	Without Pt ISD	Experimental <sup>d</sup>	Theoretical <sup>e</sup>
RM 8012	27.6 ± 2.1	0 %	27.9 ± 1.9	27.2 ± 2.0	27.9 ± 0.1	27.1 ± 0.2		
		- 20 %	27.5 ± 1.9	25.3 ± 1.7	27.3 ± 0.4	25.4 ± 0.2	- 6.8 %	- 7.2 %
		- 50 %	27.5 ± 3.0	20.0 ± 2.3	27.3 ± 0.2	20.6 ± 0.6	- 25 %	- 21 %
RM 8013	56.0 ± 0.5	0 %	55.9 ± 3.1	56.1 ± 2.9	56.2 ± 0.3	56.0 ± 0.2		
		- 20 %	55.7 ± 2.5	51.7 ± 2.3	56.1 ± 0.6	52.2 ± 0.7	- 6.9 %	- 7.2 %
		- 50 %	57.6 ± 4.5	42.8 ± 3.4	58.3 ± 1.4	42.6 ± 0.9	- 27 %	- 21 %

<sup>a</sup> Average of means of Gaussian fits to 2 or 3 individual data sets obtained by replicate runs of the same sample. Uncertainty is one standard deviation of the 2 or 3 values. Therefore, only measurement repeatability is taken into account.

<sup>b</sup> From NIST Report of Investigation. Uncertainty is expanded to a level of confidence of approximately 95 %, but includes only measurement repeatability.

<sup>c</sup> Obtained by decreasing absolute values of the extraction voltage for 50 % decrease and detector voltage for 20 % decrease.

<sup>d</sup> Relative difference between spICP-MS mean sizes with and without use of Pt ISD.

<sup>e</sup> See Fig. 1.